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#### Circulating nucleic acids in plasma or serum

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#### Abstract

Background: Nucleic acids can be found in small amounts in healthy and diseased human plasma/serum. Higher concentrations of DNA are present in the plasma of cancer patients sharing some characteristics with DNA of tumor cells. Together with decreased strand stability, the presence of specific oncogene or tumor-suppressor gene mutations, microsatellite alterations, Ig rearrangements and hypermethylation of several genes may be detected. Moreover, tumor-related mRNA has been found circulating in the plasma/serum. Conclusions: The results obtained in many different cancers have opened a new research area indicating that circulating nucleic acids might eventually be used for the development of noninvasive diagnostic, prognostic and follow-up tests for cancer. © 2001 Published by Elsevier Science B.V.

Keywords: Circulating DNA; Plasma/serum; Cancer; Diagnostic

### 1. High amounts of DNA are found in plasma / serum of cancer patients

In 1977, Leon et al. [1] reported that cancer patients harbored in their plasma higher levels of circulating DNA compared to healthy controls. Moreover, greater amounts of DNA were found in the plasma/serum of patients with metastases compared to those with localized disease. Interestingly, DNA levels decreased by up to 90% after radiotherapy, while persistently high or increasing DNA concentrations were associated with a lack of response to treatment.

After having extracted and purified the DNA from the plasma, we found in various malignancies (leu-

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kaemia, lymphoma, lung, breast and gastrointestinal tumors) that detectable amounts of circulating DNA were found only in patients with advanced malignancies bearing a large tumor cell burden [2].

## 2. DNA in the plasma/serum of cancer patients share biophysical properties with DNA of cancer cells

Increased levels of plasma DNA were found in cancer patients but it was not determined if the circulating DNA was released from activated lymphocytes reacting towards the disease or from the tumor cells themselves. Our laboratory was able to show in 1989 that this plasma DNA from cancer patients shared some biophysical properties (decreased strand stability) common to DNA of cancer cells and, hence, was of tumoral origin [2]. This approach was interesting but it required microgram

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amounts of DNA. Luckily, PCR techniques became available allowing oncogene mutation detection in minute quantities.

## 3. Tumor-related alterations in plasma / serum of patients with hematological malignancies

N-RAS mutations have been found in DNA extracted from the bone marrow of patients with myelodysplastic syndrome and acute myelogenous leukaemia (AML). These alterations have been also found in the plasma, leukocytes and bone marrow of such patients [3]. In patients with N-RAS alterations, mutant DNA was always present in plasma DNA, though sometimes absent in the DNA of peripheral blood cells or bone marrow indicating that a single bone marrow biopsy or aspiration does not necessarily contain all the malignant clones involved in the disease.

### 4. Rearranged Ig in plasma DNA as a marker for B cell malignancy

Another study on hematological disorders was done on B-cell malignancies where rearranged Ig heavy chain DNA was detected in plasma or serum samples of patients [4]. Tumor-derived clonal CDRIII DNA was found in serum or plasma in 47% of 110 patients with non-Hodgkin's lymphoma or acute B-precursor lymphoblastic leukaemia. Follow-up showed a close correlation between persisting tumor-derived plasma/serum DNA and resistant disease or early relapse.

### 5. K-RAS mutations in plasma DNA of colorectal cancer patients

Several publications [5,6] have reported the presence of *K-RAS* mutations in the circulating DNA corresponding to the mutation found in the tumor. In one of these studies, the authors have been able to follow-up a number of patients after surgery [7]. Post operatively, *K-RAS* mutations were found in the serum of five patients, two of which developed a

relapse. The plasma mutation detection predated the clinical recurrence by 1 year in one case.

### 6. K-RAS mutations in plasma DNA of pancreatic cancer patients

The *K-RAS* gene is mutated in approximately 90% of pancreatic adenocarcinomas, suggesting that the analysis of many genes would be unnecessary to detect the majority of cases. Several studies have, therefore, also been focused on the detection of this mutation in the plasma of pancreatic cancer patients [5.6].

In one study [8], plasma DNA was isolated from 21 pancreatic cancer patients and *K-RAS* alterations was detected in the plasma of 17 patients (81%). In cases where both plasma and pancreatic tissue were available, DNA mutations were similar in corresponding plasma and tissue samples. Plasma DNA alterations were found 5–14 months before clinical diagnosis in four patients who had been diagnosed as suffering from pancreatitis showing again that circulating tumor DNA may be an early event in oncogenesis. Mutant DNA was not found in the plasma of three patients with chronic pancreatitis who did not develop a carcinoma.

A follow-up study of patients with pancreatic adenocarcinoma also resulted in plasma DNA alterations being detected in a high proportion of cases in which a *K-RAS* mutation was found in tumor tissue [9]. Treatment resulted in disappearance of *K-RAS* gene mutations in plasma DNA in six of nine (67%) patients. Three patients with a persistently positive *K-RAS* gene mutation in pre- and post-treatment plasma samples showed early recurrence or a progressive disease.

### 7. Microsatellite alterations in circulating DNA of cancer patients

All tumors, however, do not have high mutation rates on easily testable hot spots. This is why several groups have looked for microsatellite alterations in the plasma/serum DNA of cancer patients. Microsatellite DNA is composed of simple repeats of unknown function. It is unstable in cancer cells and

subject to alterations, which appear as new alleles, allele expansion or loss of heterozygocity (LOH). Microsatellite DNA alterations are part of neoplastic progression and they may serve as clonal markers.

We initiated a study to detect microsatellite alterations in paired samples of plasma and tumor DNA from patients with small cell lung carcinoma compared to the same repeat sequences of normal cells from the same patients. A microsatellite alteration was present in 16/21 (76%) SCLC tumors and in 15 out of 21 (71%) corresponding plasma samples [10]. Microsatellite alterations were also found in the circulating DNA of head and neck cancer patients [11]. In this study, positive serum samples appeared to be related to patients with advanced disease, suggesting they may prove useful as prognostic factors. Microsatellite alterations have also been detected in the circulating DNA of patients suffering from a variety of malignancies: non-small cell lung [12], renal [13], bladder [14], breast [15], HNPCC and sporadic colon [16], ovarian cancers [17] and melanoma [18].

# 8. Free Epstein-Barr Virus (EBV) DNA in circulating DNA of patients with nasopharyngeal cancer

EBV sequences have been detected in the plasma of patients suffering from nasopharyngeal cancer [19]. Thus, viral DNA may serve as a tumor marker for a malignancy, which is widely distributed in Southern Asia. Quantitative analysis of circulating EBV can be related to prognosis.

#### 9. Aberrant methylation of genes in plasma/ serum of cancer patients

Epigenetic changes common in many kinds of malignancies may also be detected in the plasma or serum of cancer patients particularly promoter hypermethylation of several genes. Among these genes, p16 has been specially studied. It is known that p16 mutations are rare but promoter hypermethylation resulting in gene inactivation are found to be common in a variety of different primary tumors. New markers are available since multiple genes are now known to be somatically silenced by promoter hyper-

methylation. Twenty-two patients with non-small cell lung cancer were tested [20], searching for promoter hypermethylation of the tumor suppressor gene p16. the putative metastasis suppressor gene death-associated protein kinase, the detoxification gene glutathione S-transferase P1, and the DNA repair gene O6-methylguanine-DNA-methyltransferase. Aberrant methylation of at least one of these genes was detected in 15 of 22 (68%) NSCLC tumors but not in any paired normal lung tissue. In these primary tumors with methylation, 11 of 15 (73%) samples also had abnormal methylated DNA in the matched serum samples. Similarly, in the study of liver cancer [21], p16 methylation was found in the plasma/ serum samples of 81% (13/16) of patients presenting p16 methylation in their tumor. Hypermethylation of p16 was also reported in tumor and plasma DNA of a series of breast [22] and head and neck patients [23].

#### 10. Circulating mRNA in serum of cancer patients

RNA has also been found circulating in the plasma of normal subjects and cancer patients. Tyrosinase messenger RNA (mRNA) has been extracted from the serum of melanoma patients and subjected to RT-PCR [24]. Moreover, the presence of cell-free Epstein-Barr virus-associated RNA has also been reported in the plasma of patients with nasopharyngeal carcinoma [25]. The two human telomerase RNA subunits, telomerase RNA template (hTR) and its catalytic component (hTERT) have also been detected in the serum of patients with breast or with colorectal cancer. hTR and hTERT were undetectable in tissues and sera taken from patients with benign disease and in the sera of normal subjects [26]. Thus, it is surprising that tumor-derived mRNA is not immediately broken down in the blood stream and may be found even in patients with localized disease.

#### 11. Medical implications

From the clinical point of view, the results reported in this short review are still preliminary. Circulating DNA opens, however, a new area of research and offers the possibility of noninvasive test for cancer diagnosis. Moreover, the association between tumor-related plasma DNA and tumor stage, sometimes observed, suggests that it might potentially serve as a prognostic marker. The first practical application probably resides in the possibility to follow-up by a noninvasive test after surgery or therapy, predicting recurrence or assessing success of treatment.

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#### References

- Leon SA, Shapiro B, Sklaroff DM, Yaros MJ. Free DNA in the serum of cancer patients and the effect of therapy. Cancer Res 1977;37:646-50.
- [2] Stroun M, Anker P, Maurice P, Lyautey J, Lederrey C, Beljanski M. Neoplastic characteristics of the DNA found in the plasma of cancer patients. Oncology 1989:46:318–22.
- [3] Vasyoukhin V, Anker P, Maurice P, Lyautey J, Lederrey C, Stroun M. Point mutations of the N-ras in the blood plasma DNA of patients with myelodysplastic syndrome or acute myelogenous leukaemia. Br J Haematol 1994:86:774-9.
- [4] Frickhofen N, Muller E, Sandherr M, et al. IG heavy chain DNA is detectable in cell free blood samples of patients with B-cell neoplasia. Blood 1997;90:4953–60.
- [5] Anker P, Mulcahy H, Chen XQ, Stroun M. Detection of circulating tumor DNA in blood (plasma/serum) of cancer patients. Cancer Metastasis Rev 1999;18:65–73.
- [6] Anker P, Stroun M, editors. Circulating nucleic acids in plasma or serum. Ann. New York Acad. Sci., vol. 906, 2000: 13–30.
- [7] Ryan BM, McManus RO, Daly JS, et al. Serum mutant K-RAS in the colorectal adenoma-to-carcinoma sequence. In: Anker P, Stroun M, editors. Ann. New York Acad. Sci., vol. 906, 2000:29–30.
- [8] Mulcahy H, Lyautey J, Lederrey C, et al. A prospective study of K-ras gene mutations in the plasma of pancreatic patients. Clin Cancer Res 1998;4:271–5.
- [9] Yamada T, Nakamori S, Ohzato H, et al. Circulating DNA K-ras mutation in pancreatic adenocarcinoma. Clin Cancer Res 1998;4:1527–32.
- [10] Chen Xq, Stroun M, Magnenat JL, et al. Microsatellite alterations in plasma DNA of small cell lung cancer patients. Nat Med 1996;2:1033-5.
- [11] Nawroz H, Koch W, Anker P, Stroun M, Sidransky D.

- Microsatellite alterations in plasma DNA of head and neck cancer patients. Nat Med 1996:2:1035-7.
- [12] Sanchez-Cespedes M, Monzo M, Rosell R, Pifarré A, Calvo R, Lopez-Cabrerizo MP, et al. Detection of chromosome 3p alterations in serum DNA of non-small-cell lung cancer patients. Ann Oncol 1998;9:113–6.
- [13] Goessl C, Heicappel R, Munker R, et al. Microsatellite analysis of plasma DNA from patients with clear cell renal carcinoma. Cancer Res 1998;58:4728–32.
- [14] Utting M, Müller G, Werner W, Schubert J, Junker K. Detection of tumor genetics alterations of bladder carcinomas in body fluid depend on sample treatment before DNA isolation. In: Anker P, Stroun M, editors. Circulating nucleic acids in plasma or serum. Ann. New York Acad. Sci., vol. 906, 2000: 67–71.
- [15] Silva J, Dominguez G, Garcia JM, et al. Presence of tumor DNA in plasma of breast cancer patients: clinicopathological correlations. Cancer Res 1999:59:3251–6.
- [16] Kölble K, Ullrich OM, Pidde H, et al. Microsatellite alterations in serum DNA of patients with colorectal cancer. Lab Invest 1999:79:1145–50.
- [17] Hickey KP, Boyle DP, Jepps HM, Andrew AC, Buxton EJ, Burns PA. Molecular detection of tumor DNA in serum and peritoneal fluid from ovarian cancer patients. J Cancer 1999;80:1803–8.
- [18] Fujiwara Y, Chi DD, Wang H, et al. Plasma DNA microsatellites as tumor-specific markers and indicators of tumor progression in melanoma patients. Cancer Res 1999; 59:1567-71.
- [19] Mutirangura A, Pornthanakasem W, Theamboonlers A, et al. Epstein–Barr viral DNA in serum of patients with nasopharyngeal carcinoma. Clin Cancer Res 1998:4:665–9.
- [20] Esteller M, Sanchez-Cespedes EM, Rosell M, Sidransky D, Baylin SB, Herman JG. Detection of aberrant promoter hypermethylation of tumor suppressor genes in serum DNA from non-small cell lung cancer patients. Cancer Res 1999; 59:67–70.
- [21] Wong IH, Lo YM, Zhang J, et al. Detection of aberrant p16 methylation in the plasma and serum of liver cancer patients. Cancer Res 1999;59:71–3.
- [22] Silva JM, Dominguez G, Villanueva MJ, et al. Aberrant DNA methylation of the p16INK4a gene in plasma DNA of breast cancer patients. Br J Cancer 1999;80:1262–4.
- [23] Sanchez-Cespedes M, Esteller M, Wu L, et al. Gene promoter hypermethylation in tumors and serum of head and neck cancer patients. Cancer Res 2000;60:892-5.
- [24] Kopreski MS, Benko FA, Kwak LW, Gocke CD. Detection of tumor messenger RNA in the serum of patients with malignant melanoma. Clin Cancer Res 1999;5:1961–5.
- [25] Lo KW, Lo YM, Leung SF, Tsang YS, Chan LY, Johnson PJ, et al. Analysis of cell-free Epstein–Barr virus associated RNA in the plasma of patients with nasopharyngeal carcinoma. Clin Chem 1999;45:1292–4.
- [26] Chen XQ, Bonnefoi H, Pelte MF, et al. Telomerase RNA as a detection marker in the serum of breast cancer patients. Clin Cancer Res 2000;6:3823-6.